# Immobilization of Algicidal Bacteria for Management of Algal Blooms: A Case Study

Kathryn J. Coyne, Yanfei Wang, William C. Holland, Alan J. Kennedy and Kaytee Pokrzywinski









#### Harmful Algal Bloom (HAB) control and mitigation strategies

- > Response time
  - How quickly do they act?
  - How long do they last?
- > Specificity
  - Are they specific to a single HAB?
  - Do they have an effect on the broader community?
- > Environmental impacts
  - What is the effect on the environment?
  - How long do impacts last?









#### Competitive **Parasitic** Syneraistic metH-B<sub>12</sub> ◀ Homocysteine - NH₄ <del>◄</del> NH₄◀ Protease -DOM Fatty acid Plastid biosynthesis cycle Calvin CO2 cycle RuBisCo Mitochondrion Vibroferrin (VF) -Fe(III) ← Fe(II) ← VF-Fe(III) Nucleus cbk Glycolate HGT

Shady A. Amin et al. Microbiol. Mol. Biol. Rev. 2012; doi:10.1128/MMBR.00007-12

#### **Bacteria and Phytoplankton**

- Interactions between bacteria and phytoplankton are complex
- Bacteria may regulate algal bloom dynamics
  - Essential vitamins
  - Algicidal compounds
- Outcome of interactions are likely species specific





#### Shewanella sp. IRI-160

#### **ARTICLE IN PRESS**







Harmful Algae xxx (2004) xxx-xxx

A bacterium that inhibits the growth of *Pfiesteria piscicida* and other dinoflagellates

Clinton E. Hare, Elif Demir, Kathryn J. Coyne, S. Craig Cary, David L. Kirchman, David A. Hutchins\*

College of Marine Studies, University of Delaware, 700 Pilottown Road, Lewes, DE 19958, USA
Received 15 October 2003; received in revised form 4 January 2004; accepted 15 March 2004

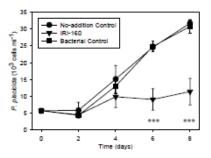
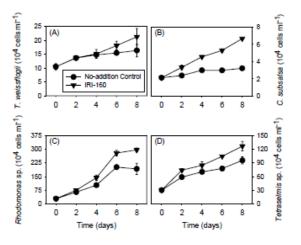


Fig. 1. Impact of bacterial strain IRI-160 ( $10^8$  cells  $ml^{-1}$ ) on P. piscicide cultures. Controls include both the addition of  $0.2 \, \mu m$  filtered sterile medium (control) and the addition of a harmless bacterium at  $10^8$  cells  $ml^{-1}$  (bacterial control, BRI-106). P. piscicide counts included only fiagellated zoospores, and encysted cells were not enumerated. Data points represent triplicate means  $\pm 1$  S.D. Significant differences between the control treatments and IRI-160 addition are indicated by \*: P < 0.05; \*\*: P < 0.01; \*\*\*: P < 0.001.



- Isolated from Delaware's inland bays and broadly distributed along the US East Coast
- Inhibits growth of a broad range of dinoflagellates, including Karenia brevis
- Stimulates the growth of other phytoplankton species





## Algicide IRI-160AA: Bacteria-free exudate from *Shewanella* sp. IRI-160

At the application rate required to control dinoflagellate growth:

- No negative effects on other phytoplankton or protists (Hare et al. 2005, Pokrzywinski et al. 2012, Tilney et al. 2017)
- No negative effects on copepods or different life stages of crabs or oysters (Simons et al. 2021)
- No evidence of primary stress response in juvenile finfish (Simons et al. 2025)









#### **Transition to Management: Application Strategies**

- 1. Dispersal of large quantities of bacteria
  - May raise concerns about biosafety
  - May dissipate quickly
- 2. Repeated dosing of IRI-160AA
  - Labor intensive
  - May dissipate quickly

#### > Solution: In situ "Bioreactor"

- Algicide produced where needed
- Limited release of bacteria
- Can be retrieved when no longer needed



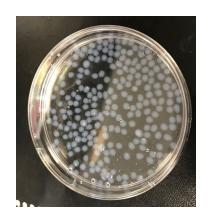


## Transition to Management: Immobilized *Shewanella* for targeted deployment

Shewanella sp. IRI-160 immobilized in alginate beads

- Biomedical and food technology industries
- Easy to prepare and store
- Can be deployed in mesh bags
- Alginate gel is biodegradable: little impact on environment

"Environmentally friendly" approach to control harmful dinoflagellate blooms

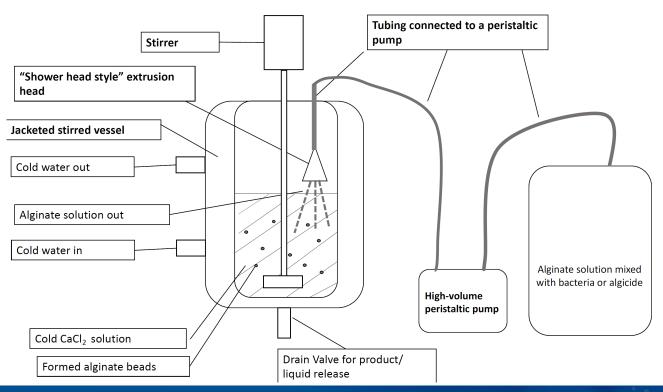


"DinoSHIELD"





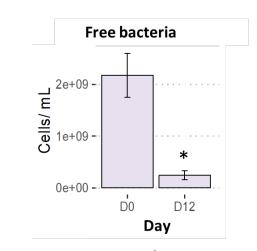
## **Alginate Hydrogels**

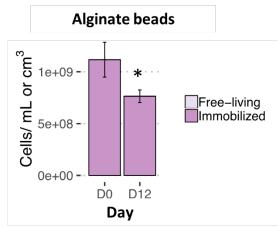


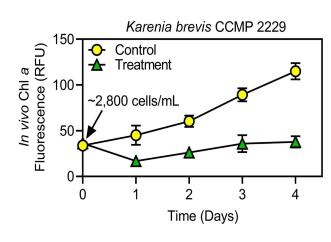




### **DinoSHIELD: Laboratory Culture Experiments**







- Protects bacteria
- Prevents dispersal
  - 99.94% of bacteria retained in matrix
     Wang and Coyne (2020)

 Effective against a broad range of dinoflagellates
 Wang and Coyne (2020)
 Wang et al. (submitted)





#### **Transition to Management: Safety Assessment**

#### 1. Environmental Impacts

What effect does DinoSHIELD have on water quality?

#### 2. Effects on Non-Target Organisms

➤ How does treatment with DinoSHIELD affect the non-target microbial community?



#### 3. Retention of Shewanella sp. IRI-160

How well does DinoSHIELD retain Shewanella in a real-world setting?





#### **Safety Assessment: Objectives**

Evaluate DinoSHIELDs within small-scale, enclosed, *in-situ* mesocosms:

- 1. Changes in water quality
- 2. Release of Shewanella bacteria
- 3. Impacts to microbial communities
  - In the absence of a bloom







#### **Methods**

- Mass produced DinoSHIELDs
- Packed in 1 μm mesh size polypropylene bags







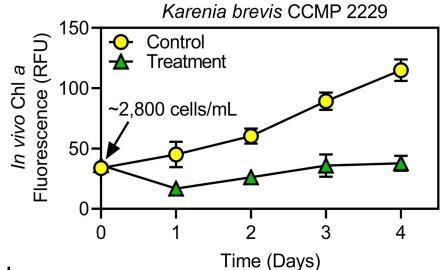






#### **Methods**

- Control (N=4): No addition
- Treatment (N=4): 3.4 L beads in
   730 L field water (v/v = 0.46%)



This rate was effective to control the growth of *Karenia brevis* in lab culture (57-67% algicidal activity)



#### **Methods**

- Water samples were collected on Day 0 before the treatment, then every day after the treatment for 6 days
- Overall photosynthetic biomass
   Eukaryotic microbial community
  - Chlorophyll a concentration
- Water quality
  - Dissolved oxygen
  - pH
  - Temperature
  - Salinity
  - Nutrients

- Eukaryotic microbial community composition and diversity
  - MicroID (diatoms, dinoflagellates, raphidophytes, ciliates)
  - 18S rRNA sequencing
- Release of Shewanella from DinoSHIELDs
  - qPCR

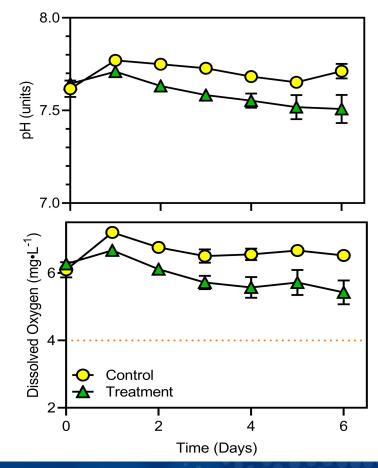




#### **Water Quality**

 No significant differences in salinity, temperature or nutrient concentrations

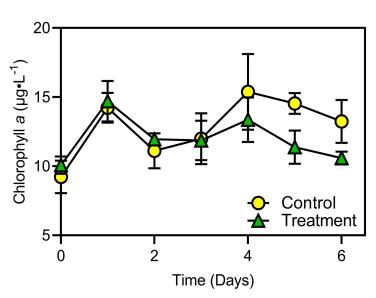
- Declining pH (~0.2) in treatment
- Decrease in dissolved oxygen
  - Still > 4 mg/L (hypoxia levels)
- Evidence of heterotrophic activity?



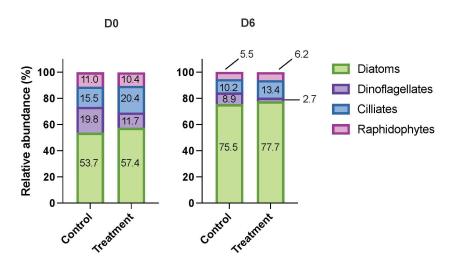




#### **Community Composition**



No significant difference in chlorophyll *a* 



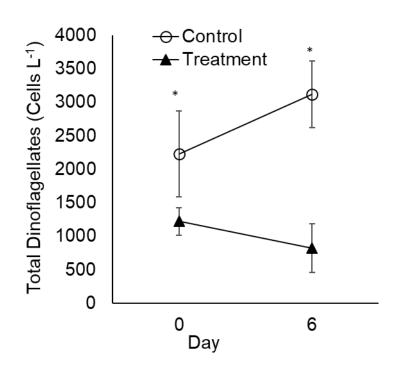
#### MicroID:

- Dominated by diatoms
- Low dinoflagellate abundance





#### **Dinoflagellate Abundance?**



Dinoflagellate abundance decreased in treatment

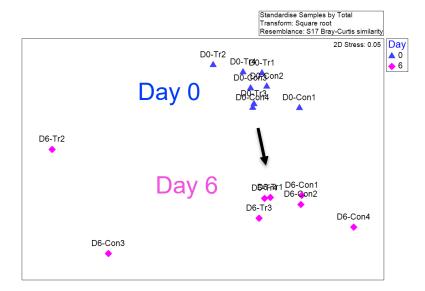


Dinoflagellate abundance was pretty low in these samples





# 18S rRNA Sequencing: Eukaryotic Microbial Community



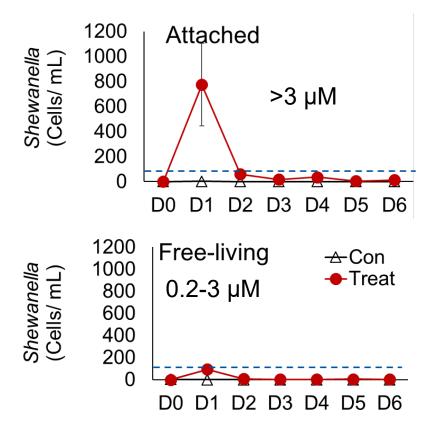
Significant changes in community structure over time

#### On Day 6:

- Dominated by diatoms
- No significant difference in community structure between controls and treatment
- ➤ Richness and diversity in treatment was significantly greater than control (p<0.05)







#### Shewanella retention

- Shewanella was released from DinoSHIELDs at very low levels
- More Shewanella attached to particles

► Total bacteria in the environment:
 > 10<sup>7</sup> cells/ mL



#### **Conclusion: Safety Assessment of DinoSHIELD**

#### 1. Environmental Impacts

- Little effect on water quality
- Slight decrease in DO and pH: Evidence for an increase in heterotrophic activity in response to DinoSHIELD

#### 2. Effects on Non-Target Organisms

- ➤ No significant effect of DinoSHIELD on community composition
- Significantly greater diversity and species richness in treatments

#### 3. Retention of Shewanella sp. IRI-160

Transient increase in Shewanella abundance, with most associated with particles >3 μm



### **Transition to Management: Future Work**

- Complete laboratory experiments to address requirements for permitting through the Federal Food, Drug, and Cosmetic Act (FDCA)
- 2. Conduct field demonstration in small, red-tide impaired embayment on southwest FL Gulf Coast (<1 acre)
- 3. Monitoring to examine changes to water quality and the microbial community after DinoSHIELD is removed from the system.





#### Acknowledgements

This project was funded by the Prevention, Control and Mitigation of Harmful Algal Blooms program at NOAA National Centers for Coastal Ocean Research (Grant# NA20NOS4780185)

Many thanks to students, staff and postdocs who participated in this project:

- Chris Holland (NCCOS)
- Alexandria Hounshell (NCCOS)
- Steve Kibler (NCCOS)
- Lonnie Gonsalves (NOAA)
- David Kidwell (NOAA)
- Tyler Harman (NCCOS)

- Mark Vandersea (NCCOS)
- Ana Rial (NCCOS)
- Lynn Wilking (NCCOS)
- Kari St. Laurent (NCCOS)
- Ashvin Iresh Fernando (ERDC)











