

# Evaluation of CRISPR/cas9 disease-resistant tomatoes for the prevention of *Salmonella* colonization

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## INTRODUCTION

- **Salmonellosis is a common food-related illness in the US** caused by several *Salmonella* strains.
- Tomato fruit can be contaminated upon *Salmonella* exposure to the plant's roots, stems, and flowers. **31 cultivars tested in FI have shown internal colonization.** <sup>1,2</sup>
- **FL8000 tomato variety was CRISPR/cas9 mutated for increased resistance** by knocking out key SAR down-regulators.
- **The CRISPR mutated tomatoes are not regulated as a genetically engineered crop.** <sup>8</sup>
- The **FL8000 mutants will be used to study whether *Salmonella enterica* Typhimurium 14028 can successfully colonize and persist in the mutated plants compared to their wild-type.**
- ***Salmonella* presence will be determined by extracting bacteria from the stem, petiole, roots, and rhizosphere.**
- Results from the study will provide knowledge to **improve food security and public health, pest and disease management, and sustainability and the environment.**
- Additionally, endophytic and rhizobial microbiomes will be studied.

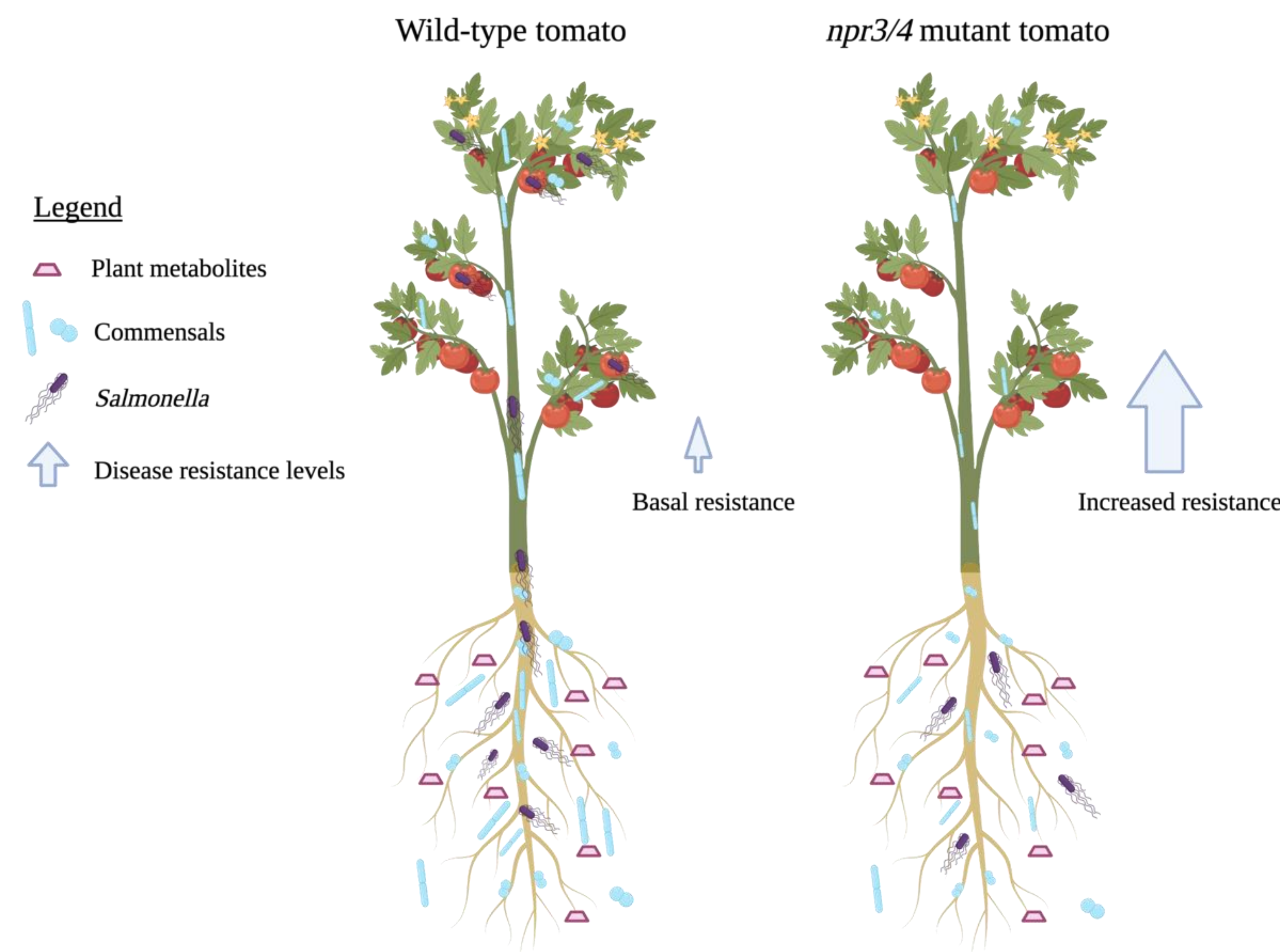
## SIGNIFICANCE

- PROS**
- Not a transgenic process = USDA doesn't regulate
  - Built-in resistance is good for plant health and food safety
  - Reduce broad-spectrum pesticides
  - Promote soil diversity
  - This CRISPR mutation can be used in other crops
  - Can be advantageous for small farmers and organic growers

- CONS**
- Not studied in the field yet
  - May have neg. consequences for commensals
  - May have neg. results in plant yield and health in the field

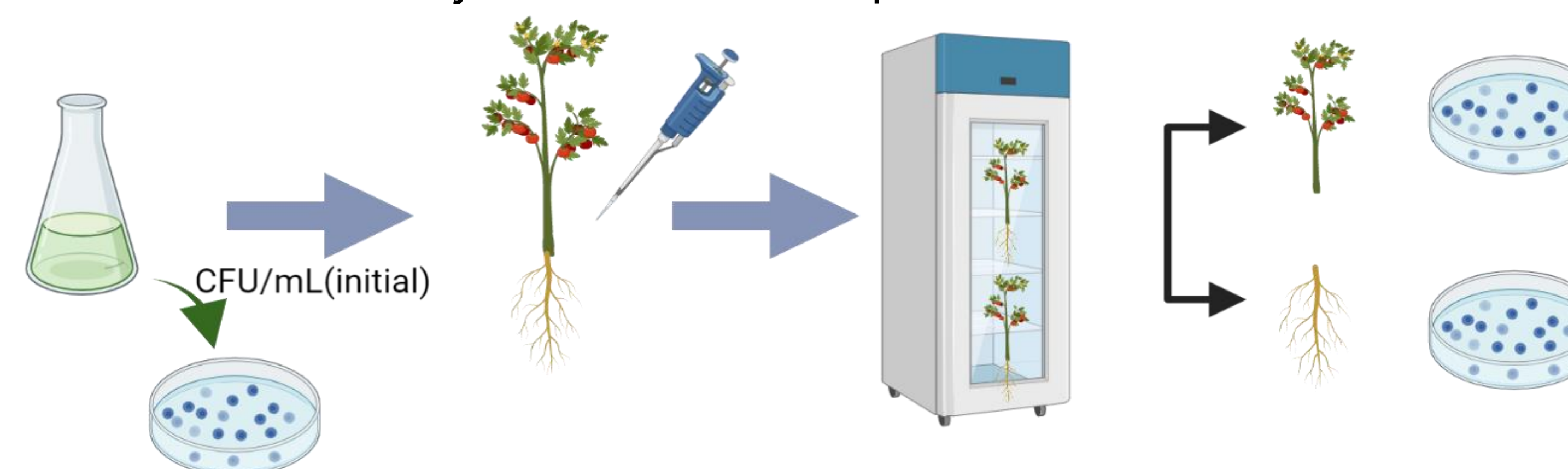
## HYPOTHESIS

CRISPR/Cas9 mutated tomato for increased plant defenses will detect MAMPs and prevent bacterial colonization and persistence compared to the unmutated wild-type parent



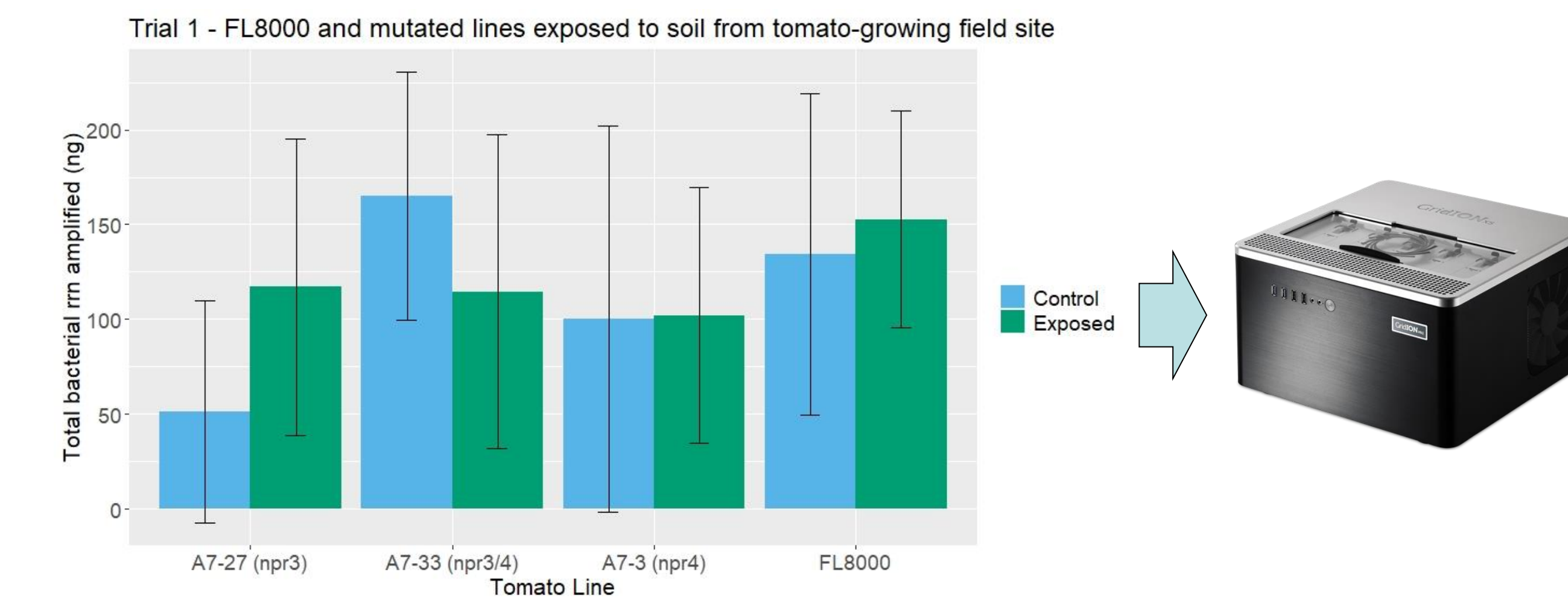
## METHODS

1. Trial 1: FL8000 and mutated lines A7-27(*npr3*), A7-3(*npr4*), A7-33(*npr3/4*) transferred to soil collected from a tomato field site in Balm, FI.
  1. Bacterial DNA extractions from apoplast to sequence endophytes.
  2. Compare endophytes in FL8000 wild-type tomato and mutated lines for differential uptake.
2. Trial 2: FL8000 and the mutated lines inoculated with GFP-*Salmonella* to determine differential uptake and colonization.
  1. Inoculations in the roots and by clippings at different concentrations.
  2. Apoplast and roots harvested. Initial and final CFU/mL determined by agar plating.
  3. Visualizations by confocal microscope



## PROGRESS AND RESULTS

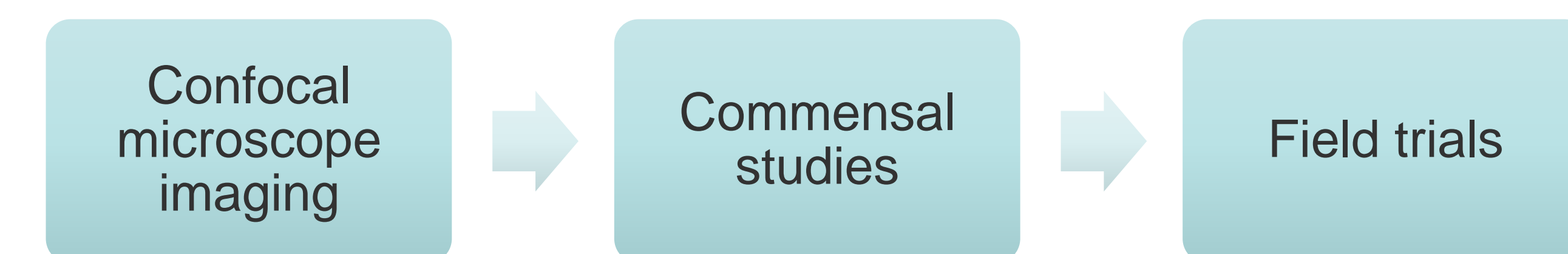
Trial 1 – pending sequencing results



Trial 2 – pending extracted CFU/mL results

Inoculation Type	Initial CFU/mL	Final CFU/mL
Soil Drench 1	10 <sup>8</sup>	Late March '22
Soil Drench 2	10 <sup>5</sup>	
Soil Drench 3	10 <sup>1</sup>	
Clipping 1	10 <sup>8</sup>	
Clipping 2	10 <sup>5</sup>	
Clipping 3	10 <sup>1</sup>	

## Upcoming work



## REFERENCES AND ACKNOWLEDGEMENTS

Han S, Micallef SA. *Salmonella* Newport and Typhimurium Colonization of Fruit Differs from Leaves in Various Tomato Cultivars. *J Food Prot.* 2014;77(11):1844-1850. doi:10.4315/0362-028X.JFP-13-562

Marvasi M, Noel JT, George AS, et al. Ethylene signaling affects susceptibility of tomatoes to *Salmonella*. *Microb Biotechnol.* 2014;7(6):545-555. doi:10.1111/1751-7915.12130

Ding Y, Sun T, Ao K, et al. Opposite Roles of Salicylic Acid Receptors NPR1 and NPR3/NPR4 in Transcriptional Regulation of Plant Immunity. *Cell.* 2018;173(6):1454-1467.e15. doi:10.1016/j.cell.2018.03.044

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RStudio Team (2021). RStudio: Integrated Development Environment for R. RStudio, PBC, Boston, MA  
 URL <http://www.rstudio.com/>

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